

Potential biomarkers associated with malignancy in uterine mesenchymal tumors

Takuma Hayashi^{1,2,*}, Nobuo Yaegashi³, Susumu Tonegawa⁴, Ikuo Konishi^{1,5}

¹Department of Cancer Medicine, National Hospital Organization Kyoto Medical Center, 612-8555 Kyoto, Japan

²Department of Industry Creation Program, START-prigram, Japan Science and Technology Agency (JST), 102-8666 Tokyo, Japan

³Department of Obstetrics and Gynecology, Tohoku University Graduate School of Medicine, 980-8575 Miyagi, Japan

⁴Department of Biology, Massachusetts Institute of Technology, Cambridge, MA 02139, USA

⁵Department of Obstetrics and Gynecology, Tohoku University Graduate School of Medicine, 606-8303 Kyoto, Japan

*Correspondence: yoyoyo224@hotmail.com (Takuma Hayashi)

DOI:10.31083/j.ejgo4205125

This is an open access article under the CC BY 4.0 license (<https://creativecommons.org/licenses/by/4.0/>).

Submitted: 18 June 2021 Revised: 16 July 2021 Accepted: 20 July 2021 Published: 15 October 2021

Uterine mesenchymal tumours, including uterine leiomyosarcoma (uLMS), are gynaecologic tumours difficult to treat clinically. To our knowledge, no standard clinical treatment for uLMS has been established in any clinical practice guideline. The primary reason for the lack of established treatment is the difficulty of conducting clinical trials because of the low incidence of uterine sarcoma. In many patients, uterine sarcoma develops within the body of the uterus. The development of sarcomas outside the body of the uterus (i.e., vagina, vulva, ovaries, etc.) is rare. A clinical study of uterine sarcoma revealed that the incidence of uterine sarcoma was 8% of malignancies that developed in the body of the uterus [1]. Uterine sarcoma is classified into carcinosarcoma, a mixed epithelial and mesenchymal tumour, and mesenchymal tumor (uLMS, endometrial stromal sarcoma, adenosarcoma, etc.). Approximately 50% of all uterine sarcomas are carcinosarcomas, and most of the other 50% are uLMSs, adenosarcomas and endometrial stromal sarcomas (ESS). Based on the results of clinical studies so far, uterine sarcomas are roughly classified into three. In clinical practice, cancer sarcoma (46%), leiomyosarcoma (36%) and ESS (13%) are found in descending order of frequency of uterine sarcoma [2, 3]. The peak age of onset is approximately 50 years for uLMS and ESS, whereas that for carcinosarcoma is ≥ 60 years [2–4]. The 50% survival for ESS is approximately 76 months, whereas that for carcinosarcoma or uLMS is approximately 27 and 30 months, respectively. Due to the low incidence of uterine sarcoma and the subtle differences in the same histology, the molecular pathological diagnosis of uterine sarcoma is often difficult in clinical practice. The clinical treatment strategy and prognosis of uterine sarcoma are determined based on the molecular pathological diagnosis of the resected uterine sarcoma tissue. Therefore, gynaecologists, radiologists and pathologists need to share molecular pathological information to determine the clinical, surgical and pathological diagnosis of uterine sarcoma.

Uterine carcinosarcoma is one of the mixed epithelial-mesenchymal tumours. The epithelial component of uterine carcinosarcoma is carcinoma, and the nonepithelial component is sarcoma. From the results of clinical research so far, single-cell origin, i.e., changes to cancer stem epithelial cells to mesenchymal cells, is considered the origin of uterine carcinosarcoma. In many patients, uterine carcinosarcoma forms a polyp-like ridge that protrudes into the uterine lumen, visible to the naked eye. Clinical studies have proposed three theories for tissue formation in uterine sarcoma, namely combination, collision and compositional tumour theory. Single-cell analysis reveals that most uterine carcinosarcoma is formed by single-cell-derived malignant tumour cells. Furthermore, it has been shown that in the process of uterine carcinosarcoma development, malignant tumour cells derived from single cells differentiate into tissues showing epithelial- and stromal-like morphologies [5]. Clinicopathological analysis reveals that uterine carcinosarcoma has more biological properties than sarcoma because of risk factors and multiple lymphoid metastases in uterine carcinosarcoma. Therefore, clinical treatment for uterine carcinosarcoma is performed according to clinical practice guidelines for high-grade uterine endometrial cancer. The 50% survival time (mOS) of patients with uterine carcinosarcoma is approximately 28 months. In other words, uterine carcinosarcoma is a poor prognosis disease. Thus, further clinical trials are needed to establish effective chemotherapy for uterine carcinosarcoma. If the uterine sarcoma component does not show a differentiation tendency, it is called homologous. The other hand, if it shows differentiation into mesenchymal tissue that does not naturally exist in the uterus, such as striated muscle, cartilage and bone, an uterine sarcoma is called heterologous. The incidence rate of uterine adenosarcoma is rare: <1% of all malignant tumours of the uterus. Therefore, uterine adenosarcoma is a malignant mixed tumour comprising benign epithelial and sarcoma components (mostly low-grade ESS). In uterine adenosarcoma, the prognosis in sar-

comatous overgrowth, in which the sarcoma component accounts for >25% of the tumour, is poor. The age of uterine adenocarcinoma onset is younger than that of carcinosarcoma onset.

For the molecular pathological diagnosis of uLMS, the surgical pathological diagnostic criteria proposed by Hendrickson and Kempson's clinical group are widely used [6]. As a surgical pathological diagnosis for uLMS, nuclear division (index), cell atypia and coagulative tumour necrosis are comprehensively evaluated. For uLMS, simple abdominal hysterectomy and bilateral adnexectomy are performed as the basic initial treatment for the excised possible cases. In uLMS, the incidence of lymphatic metastases is very low. Therefore, there is no clear evidence that extended surgery or surgical treatment of lymph node dissection improves the prognosis of patients with uLMS. Postoperative chemotherapy may be given to prevent recurrence of uLMS, but current clinical trials have not shown the efficacy of postoperative chemotherapy for uLMS. Radiation therapy for uLMS does not suppress recurrence [7].

Previously, endometrial stromal sarcoma was classified as low-grade endometrial stromal sarcoma (LG-ESS) and high-grade endometrial stromal sarcoma (HG-ESS). However, since HG-ESS not always similar to the endometrial stroma, according to the latest World Health Organisation classification, HG-ESS is classified as undifferentiated endometrial sarcoma (UES) [8]. LG-ESS of the prognosis is good, and a 50% survival period (mOS) is reportedly 76 months. Alternatively, the prognosis of UES is poor. As a treatment method for ESS, surgical treatment by simple abdominal hysterectomy and bilateral adnexectomy similar to the treatment method for uLMS is performed. However, since pelvic and para-aortic lymph node metastases were found at a rate of 9%–33% in LG-ESS and 15%–18% in UES, lymph node dissection is required for some patients with LG-ESS or UES [9, 10]. Since there are many cases in which estrogen or progesterone or both receptors are expressed in LG-ESS, hormone therapy is effective as a treatment method for LG-ESS [11]. Therefore, long-term survival of patients is recognised by hormone therapy for recurrent LG-ESS [11]. In the results of clinical trials conducted so far, the efficacy of radiation therapy and chemotherapy for ESS, regardless of malignancy, has not been confirmed.

The proteasome is a proteolytic enzyme complex consisting of multiple subunits that degrades ubiquitinated proteins in eukaryotic cells and plays a central role in proteolytic degradation. Stimulation of interferon-gamma (IFN- γ) induces the expression of beta subunits and constitutes the immunoproteasome, which regulates gene expression and cell proliferation by controlling the degradation of intracellular proteins. The expression of the major histocompatibility complex-linked low molecular mass polypeptide 2/ $\beta 1i$ (*LMP2/ $\beta 1i$*) subunit, which is increased by treatment with IFN- γ , amplifies specific endopeptidase activities of the immunoproteasome. Reports demonstrated that uLMS spon-

taneously developed after six months of age in *Lmp2/ $\beta 1i$* -deficient (mouse) female mice [12–14]. Studies have shown that the prevalence of uLMS in *Lmp2/ $\beta 1i$* -deficient mice is approximately 37% at 12 months of age [12–14]. Hematogenous metastases were also found in the *Lmp2/ $\beta 1i$* -deficient female mouse [12–14]. The incidence of other malignancies (i.e., hepatocellular carcinoma, etc.) in *Lmp2/ $\beta 1i$* -deficient mice has been reported to be 1% or less [12–14]. Therefore, in clinical research by a collaboration of medical institutions, the expression status of *LMP2/ $\beta 1i$* (Human) was examined in 74 cases with normal myometrium, uterine leiomyoma, uLMS and other uterine mesenchymal tumour tissues obtained from the pathological file by immunohistochemical (IHC) staining using an anti-human *LMP2/ $\beta 1i$* monoclonal antibody [15, 16]. As a result, the expression level of *LMP2/ $\beta 1i$* was significantly explicitly reduced in the uLMS tissues compared with those in the uterine leiomyoma and normal myometrium tissues. Therefore, it may be easy to distinguish between uLMS and uterine leiomyoma depending on the expression status of *LMP2/ $\beta 1i$* , even in cases where differential diagnosis is difficult by the surgical, pathological diagnosis method currently performed in clinical practice [15].

Based on the markedly reduced expression level of *LMP2/ $\beta 1i$* , candidate factors as biomarkers specifically expressed in uLMS have been sought using genome-wide experimental methods with human-extracted tissues. As a result, CAVEOLIN 1, CYCLIN B, CYCLIN E, Ki-67/MIB1 and *LMP2/ $\beta 1i$* were identified as biomarker candidate factors expressed explicitly in uLMS. The differential diagnostic method with IHC staining using a combination of several monoclonal antibodies to *LMP2/ $\beta 1i$* and other candidate cellular factors such as CAVEOLIN 1, CYCLIN B, CYCLIN E, Ki-67/MIB1 and 5'-nucleotidase domain containing two has been investigated for uterine mesenchymal tumours, including uLMS (Table 1) [17, 18]. Over-expression of cyclin E correlates with tumorigenesis, it is involved in various malignant tumours, including breast, colon, bladder, skin and lung cancer [19]. Recent reports have demonstrated that cyclin E-deficient cells actively proliferate in conditions of continuous cell cycling but are unable to re-enter the cell cycle from the G₁ phase to the S phase and are resistant to chemical-induced oncogenic transformation [20–22]. Cyclin E, a regulator of the cell cycle and Ki-67/MIB1, which is used as a diagnostic biomarker in proliferating cancer or malignant tumour cells, affect the behaviour of human breast cancer cells and uLMS [22–24]. Clinical studies suggested that patients with uLMS with high expression levels of cyclin E and Ki-67/MIB1 have a poor prognosis. In other words, the expression status of cyclin E and Ki-67/MIB1 is considered to correlate with the uLMS malignancy.

Mediator complex subunit 12 (MED12) is a transcriptional mediator of RNA polymerase II transcription. Previous research has shown that mutations in MED12 are found in both primary and metastatic uLMSs, including uLMS

Table 1. Differential expressions of SMA, Caveolin1, Cyclin B, Cyclin E, LMP2, NT5DC2 and Ki-67 in human uterine mesenchymal tumours and uterine LANT-like tumour.

Mesenchymal tumor types	Age years	n	Protein expression*						
			SMA	CAV1	CCNB	CCNE	LMP2	NT5DC2	Ki-67
Normal	30s–80s	74	+++	–	–	–	+++	–	–
Leiomyoma		40	+++	++	–/+	–/(+)	+++	–/+	+/-
(Ordinally leiomyoma)	30s–80s	(30)	+++	++	–/+	–	+++	–/+	+/-
(Cellular leiomyoma)		(10)	++	++	–/+	–/(+)	++	–/+	+/-
STUMP	40s–60s	12	++	++	–	–/+	–/+	–/+	+/-+++
Bizarre leiomyoma	40s–50s	4	++	++	–/+	+	Focal+	+	+
Leiomyosarcoma	30s–80s	54	–/+	+	++	+++	–/+	++	++/+++
U.LANT*-like tumour	40s	1	++	+	NA	++	–	NA	–

Staining score of expression of SMA, CAV1 (Caveolin 1), CCNB (Cyclin B), CCNE (Cyclin E), LMP2 (low molecular protein 2), NT5DC2 (5'-Nucleotidase Domain Containing 2) and Ki-67 from results of IHC experiments. Protein expression; estimated-protein expressions by immunoblot analysis, immunohistochemistry (IHC) and/or RT-PCR (quantitative-PCR), –/+; partially positive (5% to 10% of cells stained), Focal+; Focal-positive (focal or sporadic staining with less than 5% of cells stained), ++; staining with 5% or more, less than 90% of cells stained, +++; diffuse-positive (homogeneous distribution with more than 90% of cells stained), –; negative (no stained cells). U.LANT-like tumour; uterine leiomyomatoid angiomatous neuroendocrine tumour-like tumour, LMP2 (Ref. [17, 25, 26]), cyclin E (Ref. [17, 26]), caveolin1 (Ref. [27]), NT5DC2 (Ref. [28]), Ki-67 (Ref. [25, 29]). STUMP (Smooth muscle tumor of uncertain malignant potential) (Ref. [30]). Cyclin E, LMP2, Caveolin1 are potential biomarker for human uterine mesenchymal tumors. LANT*, leiomyomatoid angiomatous neuroendocrin tumour (LANT) is described as a dimorphic neurosecretory tumor with a leiomyomatous vascular component (Ref. [31]).

[29]. Protein blotting examinations revealed the expression of MED12 protein in all uterine leiomyomas and uLMS, irrespective of MED12 exon 2 mutation status or histological grade [29]. These findings suggest that MED12 has transforming roles in a broad range of smooth muscle neoplasia in addition to uterine mesenchymal tumours. As a health and welfare activity of national and international governments, uterine cancer screening (including uterine leiomyoma with high incidence regardless of race) is recommended for women aged ≥ 20 years. Micro-RNAs are short nucleic acids that regulate gene expression and protein activity in the body. About 2500 types of micro-RNAs are present in the blood. In recent years, it has been clarified that the type and amount of micro-RNAs contained in the blood fluctuate depending on the onset of cancer or malignant tumour. Using the biological properties of micro-RNAs, diagnostic techniques for cancer and malignant tumours have been investigated. Also, for preoperative uterine mesenchymal tumour screening, the usefulness of serum micro-RNA as liquid biopsy samples with high diagnostic performance has been investigated. However, because uterine mesenchymal tumours are a mixture of tumours of various morphologies and pathologies, many problems must be resolved before the results of the clinical studies can be applied to clinical practice.

Currently approved treatments for other malignancies and cancers have not been shown to be effective against uterine mesenchymal tumours, including uLMS. In international clinical research, current clinical evidence has been weighing to establish various preoperative and postoperative surgical pathological diagnostic or clinical treatment methods or both for uLMS at an early stage. Based on data obtained from clinical studies, the administration of adjuvant chemotherapy seems to show no improvement in progression-free survival

or overall survival of patients with uLMS at the early stage. Due to the low prevalence of uterine mesenchymal malignancies compared to other malignant tumours, large prospective, randomised, multi-institutional studies are required to assess better the value of different adjuvant strategies for clinical treatment against uLMS. Innovative targeted therapies must be tested by a large cohort to improve outcomes in patients with uterine mesenchymal malignancies such as uLMS, which remain inadequately effective. Pathogenic variants, including impaired expression of *LMP2/β1i* or MED12 oncogenic mutations or both, may be one of the risk factors for developing human uLMS, as are *Lmp2/β1i*-deficient mice. Thus, combining defective *LMP2/β1i* expression or MED12 oncogenic mutations or both with other functional candidates may be useful as a novel diagnostic biomarker to distinguish human uLMS from other mesenchymal tumours. To our knowledge, no effective therapy has been established for unresectable human uLMS; therefore, gene therapy with *LMP2/β1i* expression vectors might be a new treatment for human uLMS that exhibits a defect in *LMP2/β1i* expression.

ULMS is a malignant mesenchymal tumour with a poor prognosis because of repeated recurrence and distant metastasis. Preoperative diagnosis of uLMS is difficult in clinical practice. Therefore, it is important to establish a simple diagnostic method using diagnostic biomarkers to distinguish uLMS from other uterine mesenchymal tumours. Currently, clinical studies are being conducted to verify the specificity and superiority of candidate cellular factors specific to uLMS as differential diagnostic biomarkers that can predict prognosis. Healthcare workers and patients hope that a simple diagnostic method would be established based on the findings of future clinical studies.

Author contributions

TH wrote the manuscript. ST and NY provided research materials and scientific information. NY and IK carefully reviewed the manuscript and commented on aspects of clinical medicine, shared information on clinical medicine.

Ethics approval and consent to participate

This study was reviewed and approved by the Central Ethics Review Board of the National Hospital Organization Headquarters in Japan (Tokyo Japan). The authors attended a 2020 educational lecture on medical ethics supervised by the Japanese government. The completion numbers of the authors are AP0000151756 AP0000151757 AP0000151769 AP000351128. This research is not clinical study, therefore consent to participate is not required.

Acknowledgment

We sincerely appreciate the generous donation of PSMB9/ β 1i-deficient breeding mice and technical comments by Luc Van Kaer, at Massachusetts Institute of Technology. We thank Isamu Ishiwata for his generous gift of the human Ut-LMS cell lines. We appreciate the technical assistance of the research staff at Harvard Medical School. We are grateful to clinical staffs at Hyogo Cancer Center for Adults for clinical materials and helpful discussion.

Funding

This clinical research was performed with the support of the following research funding: Japan Society for the Promotion of Science for TH (grant No. 19K09840), START-program Japan Science and Technology Agency (JST) for TH (grant No. STSC20001), and National Hospital Organization Multicenter clinical study for TH (grant No. 2019-Cancer in general-02). This work was supported by grants from the Ministry of Education, Culture, Science and Technology (No. 15K10709), START-project of Japan Science and Technology Agency, the Foundation for the Promotion of Cancer Research, Kanzawa Medical Research Foundation, and The Ichiro Kanehara Foundation.

Conflict of interest

The authors declare no conflict of interest.

References

- [1] Brooks SE, Zhan M, Cote T, Baquet CR. Surveillance, epidemiology, and end results analysis of 2677 cases of uterine sarcoma 1989–1999. *Gynecologic Oncology*. 2004; 93: 204–208.
- [2] Ricci S, Stone RL, Fader AN. Uterine leiomyosarcoma: Epidemiology, contemporary treatment strategies and the impact of uterine morcellation. *Gynecologic Oncology*. 2017; 145: 208–216.
- [3] Sagae S, Yamashita K, Ishioka S, Nishioka Y, Terasawa K, Mori M, *et al*. Preoperative diagnosis and treatment results in 106 patients with uterine sarcoma in Hokkaido, Japan. *Oncology*. 2004; 67: 33–39.
- [4] Akahira J, Tokunaga H, Toyoshima M, Takano T, Nagase S, Yoshinaga K, *et al*. Prognoses and prognostic factors of carcinosarcoma, endometrial stromal sarcoma and uterine leiomyosarcoma:

- a comparison with uterine endometrial adenocarcinoma. *Oncology*. 2006; 71: 333–340.
- [5] Wada H, Enomoto T, Fujita M, Yoshino K, Nakashima R, Kurachi H, *et al*. Molecular evidence that most but not all carcinosarcomas of the uterus are combination tumors. *Cancer Research*. 1997; 57: 5379–5385.
 - [6] Bell SW, Kempson RL, Hendrickson MR. Problematic Uterine Smooth Muscle Neoplasms. *American Journal of Surgical Pathology*. 1994; 18: 535–558.
 - [7] Hayashi T. Current status of chemotherapy for uterine sarcoma in Japan. *JAMA Oncology*. 2021; 7: 1–2.
 - [8] Hendrickson MR, Tavassoli FA, Kempson RL, McCluggage WG, Haller U, Kubik-Huch RA. Mesenchymal tumours and related lesions. In Tavassoli FA, Devilee P (eds.) *World Health Organization Classification of Tumours. Pathology & Genetics. Tumours of the Breast and Female Genital Organs* (pp. 233–244). Lyon: IARC Press. 2003.
 - [9] Leath CA, Huh WK, Hyde J, Cohn DE, Resnick KE, Taylor NP, *et al*. A multi-institutional review of outcomes of endometrial stromal sarcoma. *Gynecologic Oncology*. 2007; 105: 630–634.
 - [10] Riopel J, Plante M, Renaud M, Roy M, Têtu B. Lymph node metastases in low-grade endometrial stromal sarcoma. *Gynecologic Oncology*. 2005; 96: 402–406.
 - [11] Zang Y, Dong M, Zhang K, Gao C, Guo F, Wang Y, *et al*. Hormonal therapy in uterine sarcomas. *Cancer Medicine*. 2019; 8: 1339–1349.
 - [12] Hayashi T, Kodama S, Faustman DL. LMP2 expression and proteasome activity in NOD mice. *Nature Medicine*. 2000; 6: 1065–1066.
 - [13] Hayashi T, Faustman DL. Development of spontaneous uterine tumors in low molecular mass polypeptide-2 knockout mice. *Cancer Research*. 2002; 62: 24–27.
 - [14] Hayashi T, Horiuchi A, Sano K, Hiraoka N, Kasai M, Ichimura T, *et al*. Potential role of LMP2 as tumor-suppressor defines new targets for uterine leiomyosarcoma therapy. *Scientific Reports*. 2011; 1: 180.
 - [15] Hayashi T, Horiuchi A, Aburatani H, Ishiko O, Yaegashi N, Kanai Y, *et al*. Potential biomarker for human uterine leiomyosarcoma. *Journal of Clinical Medicine Research*. 2014; 6: 392–394.
 - [16] Hayashi T, Kawano M, Sano K, Ichimura T, Gur G, Yaish P, *et al*. A novel diagnostic biomarker for human uterine leiomyosarcoma: PSMB9/ β 1i. *Chinese Clinical Oncology*. 2017; 6: 22.
 - [17] Hayashi T, Horiuchi A, Aburatani H, Yaegashi N, Tonegawa S, Konishi I. A potential diagnostic biomarker: Proteasome LMP2/ β 1i-differential expression in human uterus neoplasm. 2012. Available at: <https://www.nature.com/articles/npre.2012.7082.1> (Accessed: 2 April 2012).
 - [18] Watanabe K, Hayashi T, Katsumata M, Sano K, Abiko K, Konishi I. Development of Uterine Leiomyosarcoma during Follow-up after Cesarean Section in a Woman with Uterine Leiomyoma. *Anticancer Research*. 2021; 41: 3001–3010.
 - [19] Donnellan R, Chetty R. Cyclin E in human cancers. *FASEB Journal*. 1999; 13: 773–780.
 - [20] Geng Y, Yu Q, Sicinska E, Das M, Schneider JE, Bhattacharya S, *et al*. Cyclin E ablation in the mouse. *Cell*. 2003; 114: 431–443.
 - [21] Méndez J. Cell proliferation without cyclin E-CDK2. *Cell*. 2003; 114: 398–399.
 - [22] Hayashi T, Kawano M, Ichimura T, Ida K, Ando H, Zharhary D, *et al*. Molecular Pathology and Novel Clinical Therapy for Uterine Leiomyosarcoma. *Anticancer Research*. 2016; 36: 4997–5007.
 - [23] Keyomarsi K, Tucker SL, Buchholz TA, Callister M, Ding Y, Hortobagyi GN, *et al*. Cyclin E and survival in patients with breast cancer. *New England Journal of Medicine*. 2002; 347: 1566–1575.
 - [24] Matsuda M, Ichimura T, Kasai M, Murakami M, Kawamura N, Hayashi T, *et al*. Preoperative diagnosis of usual leiomyoma, atypical leiomyoma, and leiomyosarcoma. *Sarcoma*. 2014; 2014: 498682.
 - [25] Hayashi T, Sano K, Ichimura T, Kanai Y, Zharhary D, Aburatani H, *et al*. Characteristics of Leiomyosarcoma: Induction of

- Hematogenous Metastasis by Isolated Uterine Mesenchymal Tumor Stem-like Cells. *Anticancer Research*. 2020; 40: 1255–1265.
- [26] Hayashi T, Kobayashi Y, Sano K, Horiuchi A, Konishi I. Patent: Detection of uterine leiomyosarcoma using LMP2. Application JP2007548042A events. 2019. Available at: https://upload.u-min.ac.jp/cgi-open-bin/ctr/ctr_view.cgi?recptno=R000044182 (Accessed: 4 December 2019).
- [27] Hayashi T, Ichimura T, Yaegashi N, Shiozawa T, Konishi I. Expression of CAVEOLIN 1 in uterine mesenchymal tumors: no relationship between malignancy and CAVEOLIN 1 expression. *Biochemical and Biophysical Research Communications*. 2015; 463: 982–987.
- [28] Hayashi T, Sano K, Ichimura T, Tonegawa S, Yaegashi N, Konishi I. Diagnostic biomarker candidates including NT5DC2 for human uterine mesenchymal tumors. *International Journal of Trend in Scientific Research and Development*. 2021; 5: 604–606.
- [29] Ravegnini G, Mariño-Enriquez A, Slater J, Eilers G, Wang Y, Zhu M, *et al.* MED12 mutations in leiomyosarcoma and extrauterine leiomyoma. *Modern Pathology*. 2013; 26: 743–749.
- [30] Smooth muscle tumor of uncertain malignant potential of the uterine corpus. *Female Genital Tumours WHO Classification of Tumours* (pp. 279–280). 5th edn. WHO Classification of Tumours Editorial Board. World Health Organization. 2020.
- [31] Hayashi T, Ichimura T, Kasai M, Sano K, Zharhary D, Shiozawa T, *et al.* Characterization of Leiomyomatoid Angiomatous Neuroendocrine Tumour (LANT)-like Tumour in the Myometrium with Histopathological Examination. *Anticancer Research*. 2017; 37: 1765–1772.